

## Application Note AN # 74

## Monitoring enzyme catalysis using the VERTEX 80 FT-IR spectrometer in Rapid Scan mode

### Introduction

$\alpha$ -chymotrypsin is a well characterised mammalian digestive enzyme that catalyses the hydrolytic cleavage of peptide bonds at the carboxyl side of aromatic residues. During the chymotrypsin-catalysed hydrolysis of N-succinyl-Ala-Ala-Pro-Phe-*p*-nitroanilide (Suc-AAPF-*p*NA; see figure 1), formation of the *p*-nitroaniline product can be followed spectroscopically at 410 nm while the peptide product can be monitored by FT-IR spectroscopy due to formation of a new C-terminal carboxylate group. The VERTEX 80 FT-IR spectrometer with the UltraScan™ linear air bearing scanner with True-Alignment™ technology is ideally suited for such kinetic studies, since at the fastest mirror velocity (320 kHz) more than 100 spectra at resolution 16 cm<sup>-1</sup> can be collected per second.

### Experimental

$\alpha$ -chymotrypsin was purchased from Sigma-Aldrich (St. Louis, MO; catalogue number 27270) and used without further purification. The concentration was determined by  $\epsilon_{280} = 5104 \text{ M}^{-1} \text{ cm}^{-1}$ . We chose to use the substrate N-succinyl-Ala-Ala-Pro-Phe-*p*-nitroanilide (Sigma catalogue number S7388) as it is a 'good' substrate with a tight  $K_m$  and relatively fast  $k_{cat}$  value (see reference 1). Additionally, aqueous solutions of the substrate are reasonably soluble to about 15 mM and cleavage can also be followed spectroscopically in the near-UV with  $\epsilon_{315}$  (reactant) = 14000 M<sup>-1</sup> cm<sup>-1</sup> and  $\epsilon_{410}$  (product) = 8800 M<sup>-1</sup> cm<sup>-1</sup>. The FT-IR stopped-flow instrument consists of a drive unit, a

thermostatted umbilical supply tube and an infrared cell with an integrated mixer ([TgK Scientific, Bradford on Avon, UK](#)) mounted in the sample compartment of a Bruker VERTEX 80 FT-IR spectrometer. The stopped-flow unit and the mixing cell are both contained within an anaerobic Belle Technology glove box, which allows oxygen-sensitive reactions to be performed under a nitrogen environment containing <5 ppm oxygen. The infrared transmission cell has been described previously (see

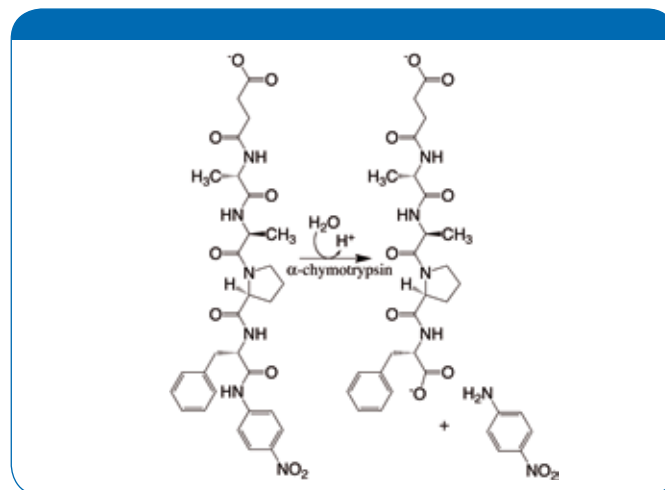


Figure 1: The  $\alpha$ -chymotrypsin-catalysed cleavage of Suc-AAPF-*p*NA yielding Suc-AAPF-COO<sup>-</sup> and *p*-nitroaniline.

reference 2). Briefly, the cell is a demountable stainless steel unit with an integrated T-mixer. It has 1.2 cm CaF<sub>2</sub> windows and the flow channels are 0.5 mm<sup>2</sup>, together creating an 8 mm diameter observation chamber with a 100 μm path length. The stopped-flow mixing time is < 10 ms and the shot volume is variable, with a value of 200 μl used in this study. A photo of the apparatus is shown.

2 mM α-chymotrypsin was mixed with an equal volume of 15 mM Suc-AAPF-pNA in 50 mM potassium phosphate/D<sub>2</sub>O, pD 8.4 at room temperature within the stopped-flow FT-IR apparatus. A narrow band MCT detector was used for speed and sensitivity. In addition, a long wave pass optical filter <1828 cm<sup>-1</sup> is required. The interferogram acquisition mode of "double-sided forward-backward" gave spectra every ~ 68 ms for ~35 s with a spectral resolution of 4 cm<sup>-1</sup> (see figure 2).

Further post measurement processing by splitting the interferograms by software enables a four fold increase in time resolution down to ~17 ms.

## Results

The C-terminal carboxylate moiety of the product is monitored as an increase in both C=O and C-O stretches at 1604 and 1322 cm<sup>-1</sup> respectively (see difference spectra in figure 3). The disappearance of the peaks at 1521 and 1344 cm<sup>-1</sup> is assigned to the NO<sub>2</sub> group. Representative reaction traces are shown in figure 4 with their colour referring to the peaks marked in the difference spectra. The difference spectra show clear isosbestic points suggestive of a single chemical reaction and reaction traces show that the reaction is completed after ~10 s.

## Acknowledgements

These experiments were performed in the Manchester Interdisciplinary Biocentre at the University of Manchester, England.

## References

1. Delmar, E.G., Largman, C., Brodrick, J.W., Geokas, M.C. (1976) *Anal. Biochem.* 99, 316
2. Thumanu, K., Cha, J., Fisher, J.F., Perrins, R., Mobashery, S., Wharton, C. (2006) *Proc. Natl. Acad. Sci. USA*, 103, 10630

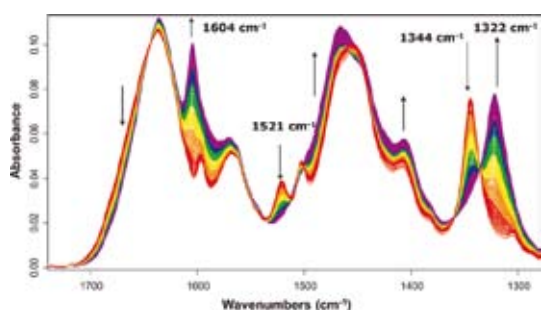


Figure 2: Progressive FT-IR spectra recorded over the time course of the reaction



Photo showing stopped flow unit in foreground and VERTEX 80 with IR cell in background

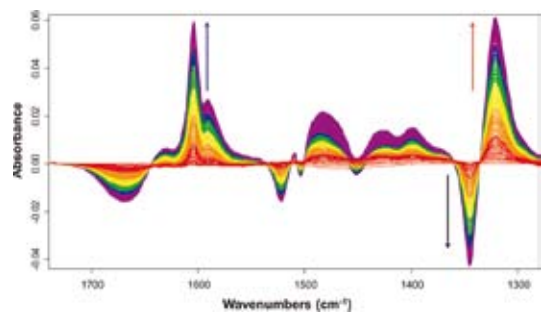


Figure 3: Difference FT-IR spectra after subtraction of the first spectrum which highlight the intensity changes

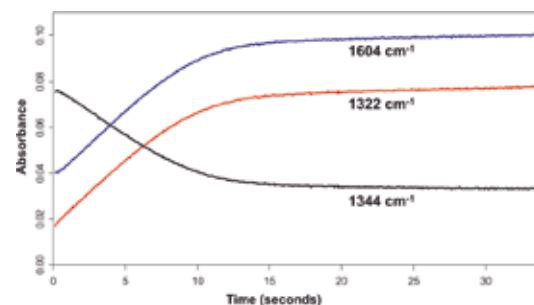


Figure 4: Peak heights of selected bands of the FT-IR spectra versus time which monitor the reaction rate.

[www.brukeroptics.com](http://www.brukeroptics.com)



**Bruker Optics Inc.**

Billerica, MA · USA  
Phone +1 (978) 439-9899  
Fax +1 (978) 663-9177  
info@brukeroptics.com

**Bruker Optik GmbH**

Ettlingen · Deutschland  
Phone +49 (7243) 504-2000  
Fax +49 (7243) 504-2050  
info@brukeroptics.de

**Bruker Optik Asia Pacific Ltd.**

Hong Kong  
Phone +852 2796-6100  
Fax +852 2796-6109  
asiapacific@brukeroptics.com.hk